

The effect of SARS-CoV-2 variant B.1.1.7 on symptomatology, re-infection and transmissibility

Mark S. Graham^{1*}, Carole H. Sudre^{1,2,3*}, Anna May⁴, Michela Antonelli¹, Benjamin Murray¹, Thomas Varsavsky¹, Kerstin Kläser¹, Liane S. Canas¹, Erika Molteni¹, Marc Modat¹, David A. Drew⁵, Long H. Nguyen⁵, Lorenzo Polidori⁴, Somesh Selvachandran⁴, Christina Hu⁴, Joan Capdevila Pujol⁴, The COVID-19 Genomics UK (COG-UK) consortium⁶⁺, Alexander Hammers¹, Andrew T. Chan⁵, Jonathan Wolf⁴, Tim D. Spector⁷, Claire J. Steves⁷⁺, Sebastien Ourselin¹⁺

1. School of Biomedical Engineering & Imaging Sciences, King's College London, London, UK
2. MRC Unit for Lifelong Health and Ageing, Department of Population Science and Experimental Medicine, University College London, UK
3. Centre for Medical Image Computing, Department of Computer Science, University College London, UK
4. Zoe Global Limited, London, UK
5. Clinical and Translational Epidemiology Unit, Massachusetts General Hospital and Harvard Medical School, Boston, MA, USA
6. <https://www.cogconsortium.uk>
7. Department of Twin Research and Genetic Epidemiology, King's College London, London, UK

* Equal contribution

+Equal contribution

+Full list of consortium names and affiliations are in the appendix

Corresponding Author:

Mark Graham, PhD

School of Biomedical Engineering and Imaging Sciences

King's College London,

Lambeth Palace Road,

SE1 7EH

mark.graham@kcl.ac.uk

Abstract

The new SARS-CoV-2 variant B.1.1.7 was identified in December 2020 in the South-East of England, and rapidly increased in frequency and geographic spread. While there is some evidence for increased transmissibility of this variant, it is not known if the new variant presents with variation in symptoms or disease course, or if previously infected individuals may become reinfected with the new variant. Using longitudinal symptom and test reports of 36,920 users of the Covid Symptom Study app testing positive for COVID-19 between 28 September and 27 December 2020, we examined the association between the regional proportion of B.1.1.7 and reported symptoms, disease course, rates of reinfection, and transmissibility. We found no evidence for changes in reported symptoms, disease severity and disease duration associated with B.1.1.7. We found a likely reinfection rate of around 0.7% (95% CI 0.6-0.8), but no evidence that this was higher compared to older strains. We found an increase in $R(t)$ by a factor of 1.35 (95% CI 1.02-1.69). Despite this, we found that regional and national lockdowns have reduced $R(t)$ below 1 in regions with very high proportions of B.1.1.7.

Introduction

In early December 2020, a phylogenetically distinct cluster of SARS-CoV-2 was genetically characterised in the South-East of England. The majority of cases had been detected in November with a small number detected as early as September¹. Genomic surveillance reveals that this new variant, termed B.1.1.7, has a number of mutations of immunologic significance and is growing rapidly in frequency and spread.²

Preliminary evidence from epidemiological studies suggests the new strain is more transmissible. Davies et. al. found the new strain is 56% (95% CI 50-74) more transmissible³ and Volz et. al. found the new strain increases the effective reproduction number $R(t)$ by a factor of 1.4-1.8⁴. There is early data to suggest B.1.1.7 increases risk of death by ~ 1.3 .⁵ However, there is much that is still unknown. Little is known about the disease course of infections due to the new variant. Early evidence suggested that the new variant does not affect rates of hospitalisation³, but it is crucial to assess whether the new variant alters the symptomatology, duration, and severity of disease. It is also important to understand whether B.1.1.7 alters the rate of asymptomatic infection and reinfection. Furthermore, early estimates of the new transmissibility of B.1.1.7 are uncertain and there is a need for additional estimates using independent data sources.

We make use of data from the COVID Symptom Study (CSS)⁶ to investigate the symptomatology, disease course, and transmissibility of the new variant. The longitudinal dataset provides symptom reports and test results from a population of over 4 million adults living in the UK using the mobile application. By combining these data with surveillance data from the Covid-19 UK Genetics Consortium (COG-UK)⁷ and a spike-gene target failure correlate in community testing data, we performed associative studies to study the symptoms, disease course, rates of reinfection, and transmissibility of the new variant.

Methods

Symptom study data

Longitudinal data were prospectively collected using the CSS app, developed by Zoe Global with input from King's College London (London, UK), the Massachusetts General Hospital (Boston, MA, USA), and Lund and Uppsala Universities (Sweden). The app guides users through a set of enrolment questions, establishing baseline demographic and health information. Users are asked to record each day whether they feel physically normal, and if not, to log any symptoms. Users are also asked to maintain a record of any COVID-19 tests, their type, and their results in the app. Users are able to record the same data on behalf of others, such as family members, to increase data coverage amongst those unlikely to use mobile applications, such as the elderly. More details about the app can be found in a study by Drew and colleagues⁶.

Genomic data

We used data released on 13 January 2020 from COG-UK to extract time-series of the percentage of daily cases that came from the B.1.1.7 lineage in Scotland, Wales, and each of the seven National Health Service (NHS) regions in England. Northern Ireland was excluded due to the low number of samples in the COG-UK dataset. These data are produced by sequencing a sample of polymerase chain reaction (PCR) tests carried out in the community. Due to the delay of approximately two weeks² between PCR and genomic sequencing, we only used data from samples taken up to 31 December to avoid censoring effects.

Additionally, we used data from Public Health England (PHE) on the probable new variant captured in community cases in England using spike gene target failure (SGTF). It has been observed that one of the spike gene mutations in B.1.1.7 causes an SGTF in the test used in three of England's large laboratories used for analysis of community cases.¹ This failure results in a marker that is sensitive to B.1.1.7, but not necessarily specific, as other circulating variants also contain the mutation leading to an SGTF. Comparison to genomic data finds that from 30 November 2020 onwards more than 96% of cases with the SGTF were from lineage B.1.1.7⁸. The proportion of SGTF cases is made available in England for each of the 316 "Lower Tier" Local Authorities. We grouped these data to each NHS region using a population-weighted average to enable integration with other data sources.

Disease symptoms and course

In order to assess whether the symptomatology of infection from B.1.1.7 differed from previous variants, we investigated the change in symptom reporting from 28 September to 27 December 2020, covering 15 complete weeks over the period when the proportion of B.1.1.7 grew most notably in London, South East and East of England. We took the symptom reports from users reporting a positive swab test (PCR or lateral flow) in this period and examined the association between the proportion of B.1.1.7 in each region and the proportion of reports per week for each

symptom, accounting for age, sex, and two seasonal environmental confounders: regional temperature and humidity, in a linear regression. Seasonal confounders were calculated each day as the average of the temperature and relative humidity at two meters above the surface, averaged across each region considered.⁹

We also examined the relation between proportion of B.1.1.7 and disease burden, measured here as the total number of different symptoms reported over a period of two weeks before and two weeks after the test, and the relation with asymptomatic infection, defined as users reporting a positive test result but no symptoms in the two weeks before or after the test. Using similar corrections for demographic and seasonal environmental confounders, we investigated the rate of self-reported hospital visits. We also investigated the proportion of individuals reporting long symptom duration using a previously published definition of continuous symptoms reported for at least 28 days.¹⁰ To avoid censoring effects, both hospitalisation and long duration analyses included symptom reports extended up to 18 January, and the long duration analysis only considered reports of positive tests up to 21 December.

Reinfection

We defined possible reinfection as the presence of two reported positive tests separated by more than 90 days with a period of reporting no symptoms for more than seven days before the second positive test. We calculated the proportion of possible reinfection among individuals reporting their first positive test before 1 October 2020 and the correlation between number of possible reinfections and number of reported positive tests. To assess whether the risk of reinfection was stronger in the presence of the new variant, we calculated the correlations between the number of possible reinfections and the proportion of B.1.1.7 cases regionally over time.

Transmissibility

Daily incidence for Scotland, Wales, and each of the seven NHS regions in England were produced from the period 1 October 2020 to 27 December 2020 using data from the CSS app and previously described methodology¹¹. These data were used to determine the number of new daily cases from both old variants and from B.1.1.7 in each region. $R(t)$ was estimated separately for the old and new variants using methods described in¹¹. We compared both multiplicative and additive differences of the new and old R values for days when the proportion of B.1.1.7 in a region was greater than 3%. While data is not available for the proportion of B.1.1.7 in January, we also computed total incidence and R from 1 October to 16 January to see the effect of national lockdown in England on these measures.

Results

Symptom study data

Table 1 shows the demographic data for the cohort studied. From 24 March to 27 December 2020, 4,327,245 participants from the UK signed up to use the app. We excluded users living in Northern Ireland due to the low number of sign-ups (38,976), 383,352 users lacking information on sex, and 2,175,979 who had not logged in the app during the period 28 September to 27 December 2020, leaving a total of 1,767,914 users. Between them, these users recorded 65,606,869 logs in the app between 28 September and 27 December. In this period, 497,989 users reported a swab test. 55,192 of these reported a positive test, and we investigated the symptom reports of 36,920 of those whose region was known and who reported as healthy on app sign-up.

		Overall		Tested		Tested positive		Signed up healthy with reporting around positive test	
		N	%	N	%	N	%	N	%
Users		1,767,914	---	497,989	---	55,192	---	40,463	
Daily reports*		65,613,697		19,154,601		1,514,244			
Age in years mean (std)		48.4 (19.3)	---	46.06 (17.8)	---	42.1 (16.8)	---	42.9 (17.0)	
	≤18	163,112	9.2	40,717	8.2	5,468	9.9	3,874	9.6
	19 - 64	1,234,259	69.8	381,900	76.7	45,149	81.8	32,878	81.2
	≥ 65	370,543	20.9	72,741	14.6	4,367	7.9	3,600	8.9
	Invalid	5,576	0.3	2,631	0.5	208	0.3	111	0.3
Sex	Female	1,046,074	59.2	315,875	63.4	34,516	62.5	24,844	61.4
	Male	720,562	40.8	181,110	36.4	20,546	37.2	15,545	38.4
	Intersex	79	<0.1	21	<0.1	3	<0.1	3	<0.1
	Prefer not to say	1,199	0.1	983	0.2	127	0.2	71	0.2
Region	South East	342,881	19.4	97,143	19.5	8,762	16.0	6,555	16.2

	East of England	196,063	11.1	57,680	11.6	5,373	9.8	4,037	10.0
	London	227,004	12.8	81,940	16.5	9,733	17.8	7,384	18.2
	Midlands	198,350	11.2	57,582	11.6	6,695	12.2	4,756	11.8
	North East and Yorkshire	156,999	8.9	42,986	9.1	5,292	9.7	3,744	9.3
	North West	123,201	7.0	45,156	9.1	6,180	11.3	4,399	10.9
	South West	186,372	10.5	46,780	9.4	3,685	6.7	2,637	6.5
	Scotland	87,263	4.9	13,793	2.8	1,589	2.9	1,049	2.6
	Wales	82,886	4.7	16,471	3.3	3,092	5.6	2,359	5.8
	Not known	165,164	9.3	38,458	7.5	4,638	8.0	3,543	8.8

Table 1. Characteristics of GB app users active in the period 28 September - 27 December 2020

* Reports logged between 28 September - 27 December. For some analyses we took further reports from an extended time period 14 September 2020 - 18 January 2021

**May be more than one test per individual as the overall number contains failed tests and unknown results

Genomic data

In the period between 27 September 2020 and 31 December 2020, 98,170 sequences were made available by COG-UK, corresponding to 4.4% of the 2,207,476 cases recorded in this period.¹² 16,224 sequences (16.5%) were variant B.1.1.7. Considering the mean of the rolling average across December, the three regions with the largest proportion of B.1.1.7 are the South East, London, and East of England. The three regions with the lowest proportion are Wales, the North East and Yorkshire, and the North West. SGTF data was made available in England on a weekly basis from 10 November 2020 to 29 December 2020. Of the 700,590 cases reported in this period, 295,404 (42.2%) caused an SGTF. Examining the COG-UK data from England in the same time period, we find 34.6% cases are B.1.1.7. The difference is in part attributable to the SGTF being a nonspecific marker of B.1.1.7: in the week from 9-15 November 81% of cases with an SGTF were B.1.1.7, while from 30 November at least 96% of cases with the SGTF were from B.1.1.7. Figure 1 shows how the proportion of the new variant changed over time in regions of the UK using COG-UK and the SGTF data.

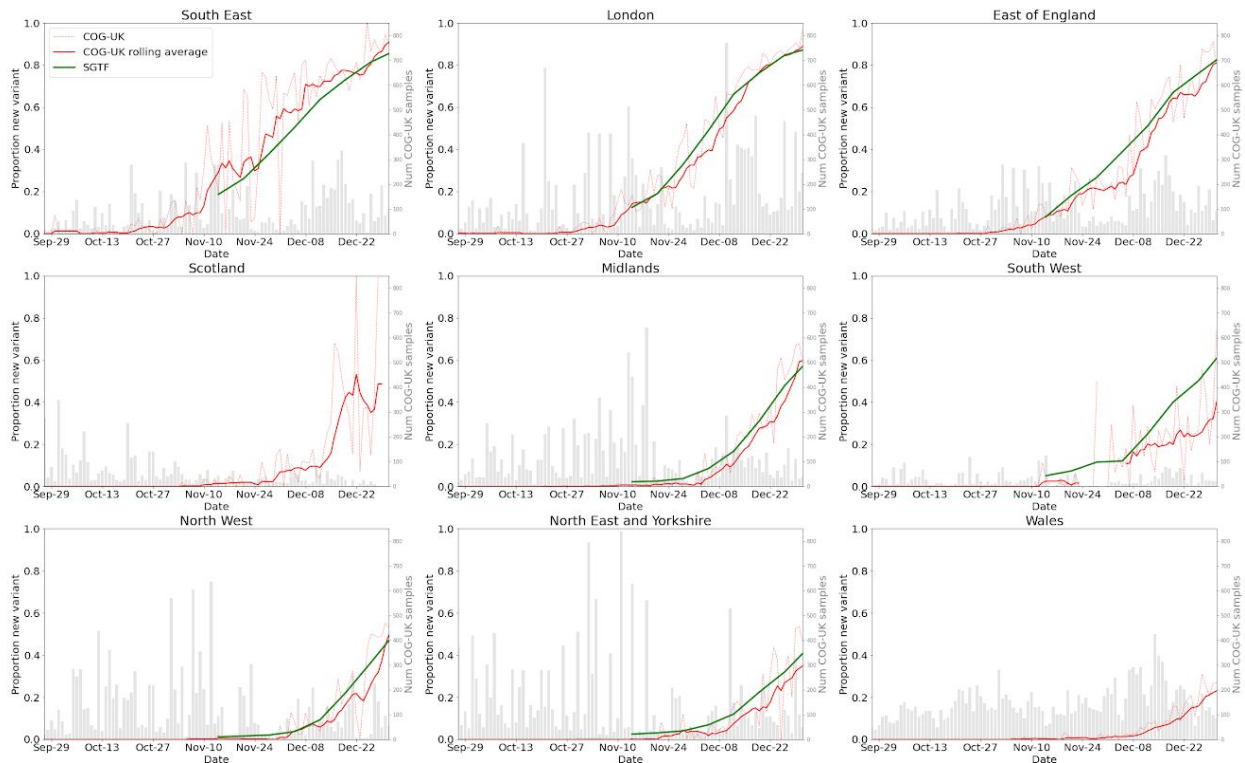


Figure 1. Presence of B.1.1.7 in each of the 7 NHS regions in England, and Scotland and Wales, as measured using genomic surveillance data (COG-UK) and SGTF data. SGTF data are not available for Scotland or Wales.

Disease symptoms and course

Figure 2 illustrates the variation of symptom occurrence over time considering a one-week window smoothed over 3 time points as a function of time, and Supplementary Figure 1 shows how these symptoms vary as a function of the proportion of B.1.1.7. These results show no change in the proportion of users reporting each symptom with the new variant.

Figure 3 shows the variation of total number of symptoms reported, the total number of asymptomatic infections, self-reported hospital visits, and symptoms of long duration over time; Supplementary Figure 2 shows how these plots vary with proportion of B.1.1.7. When correcting for mean age, sex, ambient temperature and humidity there was no evidence of an association between B.1.1.7 and either the number of symptoms reported over a 4-week window, the number of hospitalisations, long symptom duration, or proportion of asymptomatic case (Supplementary Table 1).

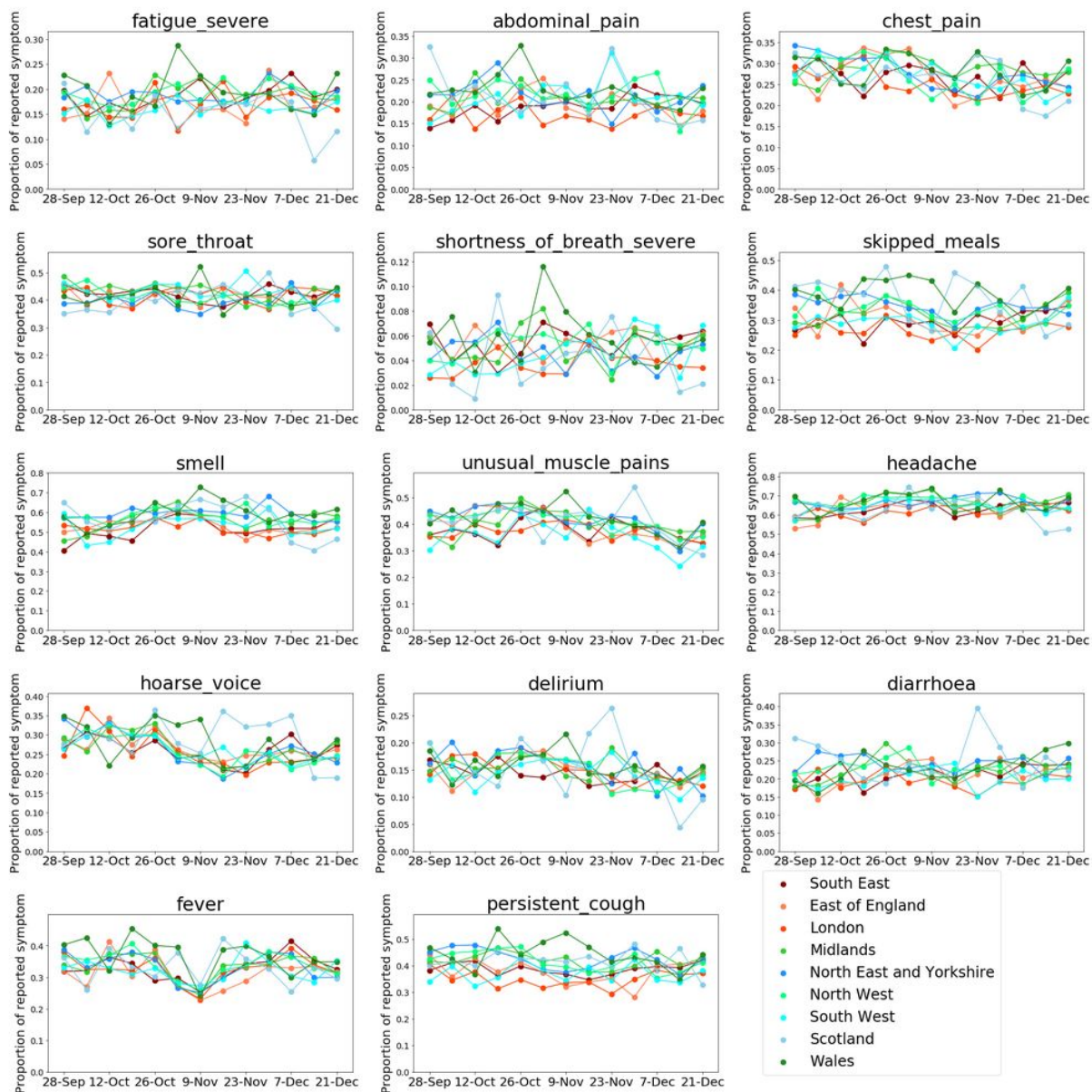


Figure 2. Regional plots of the frequency of reporting of symptoms over time for each reported symptom. Drop in fever reporting in early November was caused by a change in the question wording; this wording was subsequently reverted a week later.

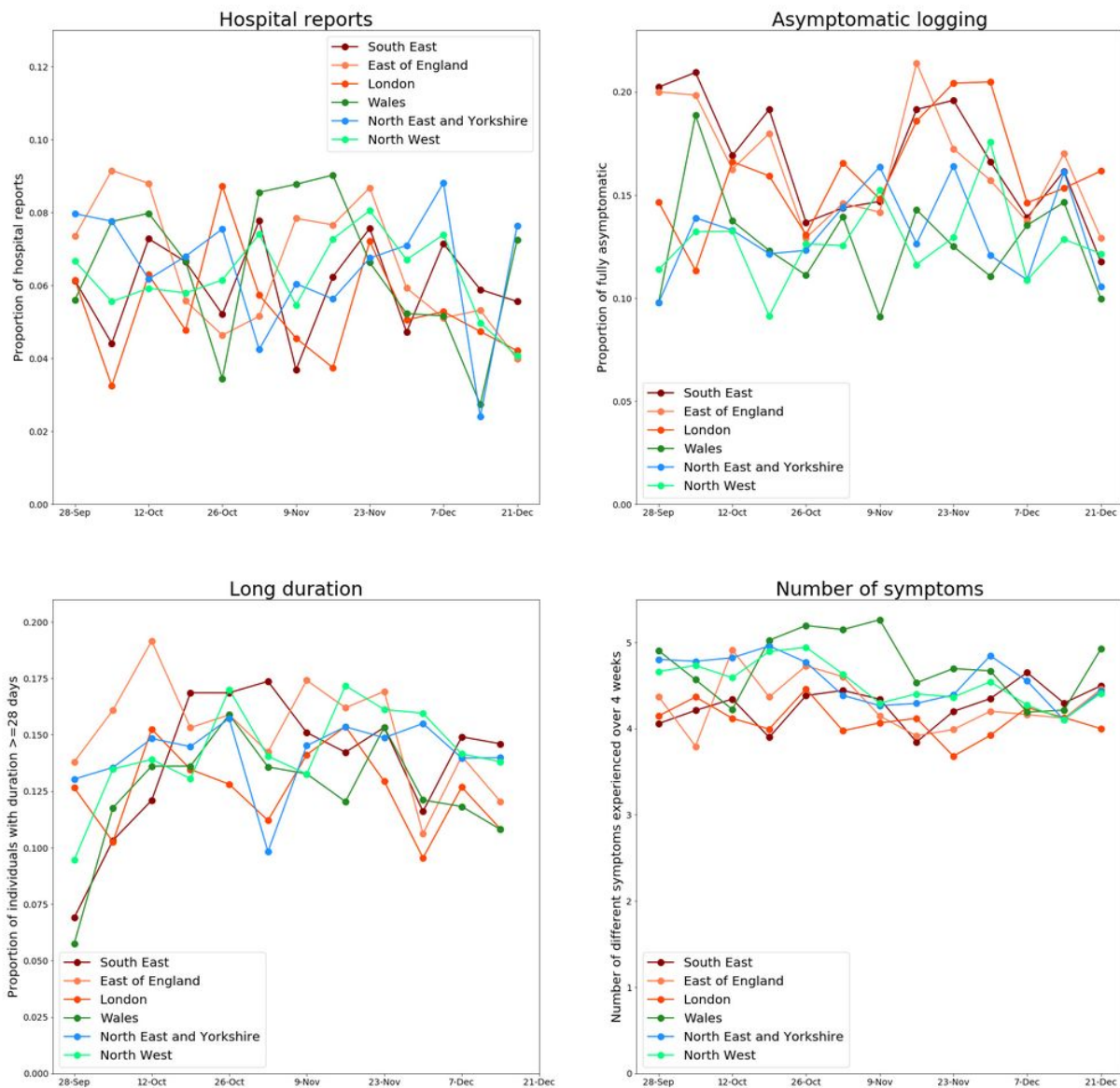


Figure 3. Regional plot of hospitalisation report, proportion of asymptomatic, report of long duration and number of experienced symptoms around test. For the study of long duration, tests are only considered up to 21 December counting reports up to 18 January 2021 to limit right censoring. Only symptomatic individuals for which duration can be ascertained are included.

Reinfection

Overall, we identified 304 individuals reporting two positive tests with more than 90 days between the two. Among these individuals, symptom reporting allowed us to identify 249 for which there is a period of at least 7 symptom-free days in between positive tests among the 36,509 individuals having reported a positive swab test before 1 October 2020 (0.7%, 95% CI 0.6-0.8). Among those 249, daily reports were available in the periods around both of the positive tests for 173. There was no difference in reinfection reporting rates across the different NHS regions ($p=0.1$). Figure 4 shows the evolution in the number of possible reinfections along

with reported positive cases (red line) and proportion of B.1.1.7 (green line). For all regions (except Scotland), reinfection occurrences were more positively correlated with the overall regional rise in cases rather than the regional rise in the new variant percentage (Number of cases:reinfection, Spearman rho 0.55 to 0.69 [$p < 0.05$] for South East, London and East of England; % new variant:reinfection, Spearman rho 0.37 to 0.55 in the same regions). Supplementary Table 2 shows the bootstrapped median values of correlation compared across the different regions and the outcome of a Mann-Whitney U test across the bootstrapped distributions.

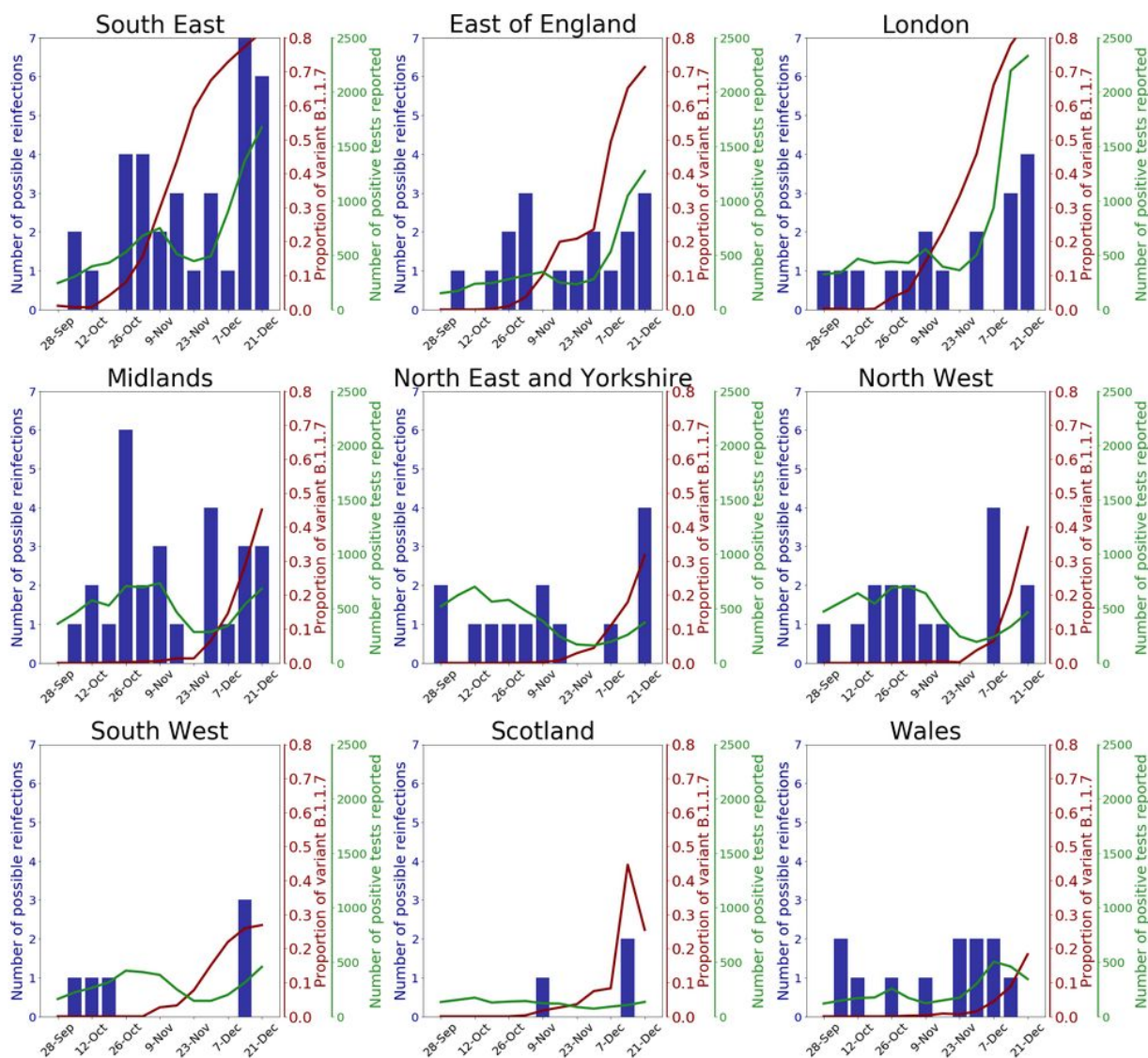


Figure 4. Number of reinfection reports by region according to week of second infection, along with the total number of positive tests reported through the app and the proportion of B.1.1.7 in circulation.

Transmissibility

Figure 5 shows incidence and $R(t)$ for the old and new variants in the three regions in England with the highest proportions of the new variant. Results consistently show the $R(t)$ of B.1.1.7 to be greater than that of other variants. The mean (95% CI) of the additive increase in R for B.1.1.7 was 0.34 (0.02-0.66), and the multiplicative increase was 1.35 (1.02-1.69). England exited its second national lockdown on 2 December, leading to a change in behaviour and $R(t)$. When considering only the period after the second lockdown ended, we find 0.28 (0.01-0.61) for the additive and 1.28 (1.02-1.61) for the multiplicative increases. Supplementary Figure 3 shows the same using the SGTF data, with analysis limited to the period after 1 December when at least 95% of all SGTF cases were B.1.1.7. These data are provided weekly, and linear interpolation was used to obtain daily estimates, leading to smoother estimates for variant-specific incidence and $R(t)$. Using these values, we find $R(t)$ of B.1.1.7 has an additive increase of 0.26 (0.15-0.37) and a multiplicative increase of 1.25 (1.17-1.34).

On 19 December 2020 London and much of the South East and East of England were placed in ‘Tier 4’ restrictions, enforcing stricter rules for social distancing and decreased human-to-human contact that stopped short of nationwide measures. On 5 January 2021 the whole of England was placed in national lockdown. Figure 6 shows overall incidence and $R(t)$ for the longer period from 1 October 2020 to 16 January 2021 in the three regions with the largest proportion of B.1.1.7. The proportion of B.1.1.7 in these regions in January is at least 80%, assuming the proportion has not decreased from the end of December. The combination of Tier 4 and national lockdown measures were able to bring $R(t)$ to ~ 0.8 in all three of these regions.

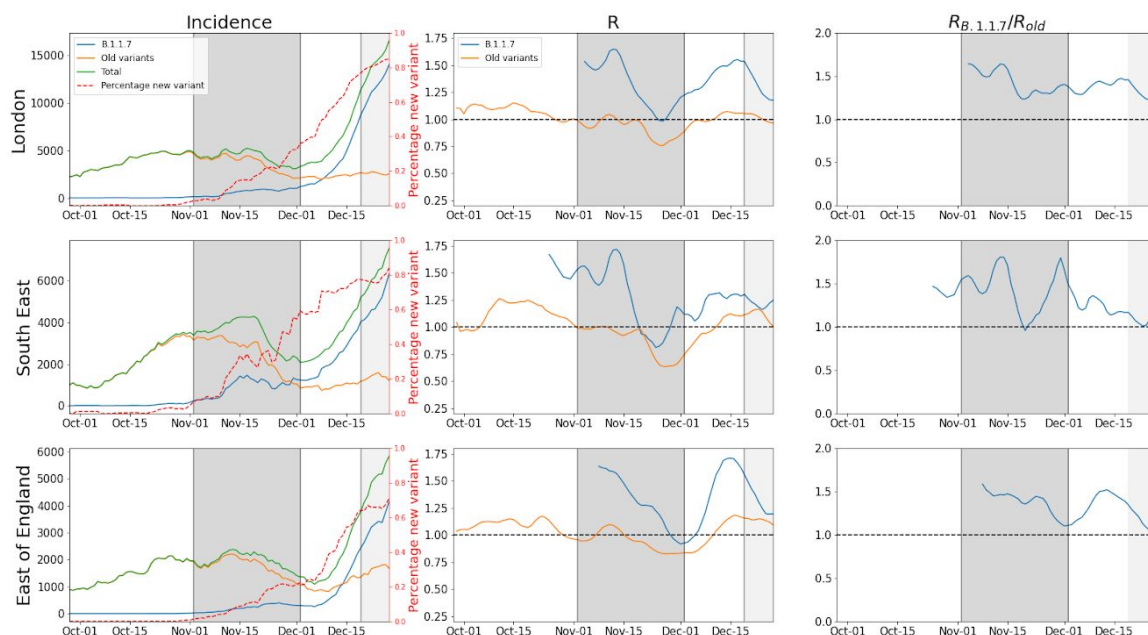


Figure 5. Incidence and $R(t)$ for the old and new variants, along with the ratio between these R values, for the three regions in England with the largest proportion of B.1.1.7. Dark grey regions indicate national lockdowns, light grey the period where London and much of the South East and East of England were placed in Tier 4 restrictions.

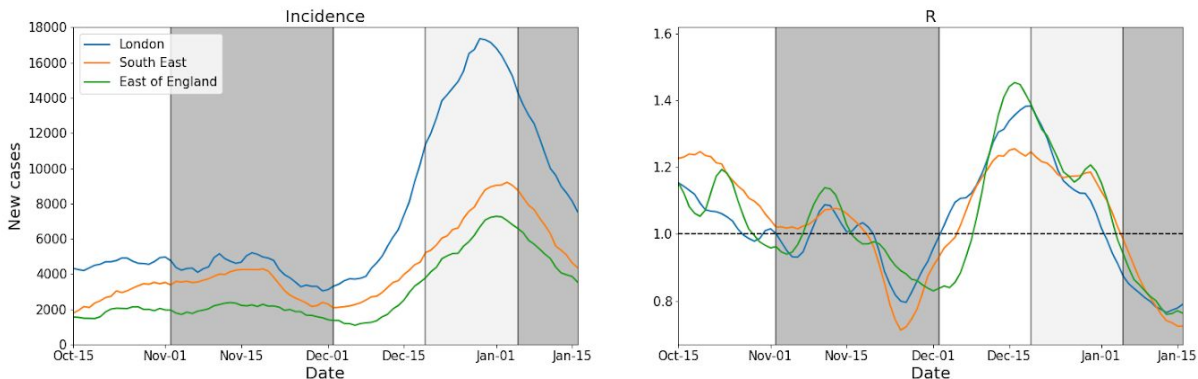


Figure 6. Total incidence and $R(t)$ for the three regions with the highest proportion of B.1.1.7 in December, extended to capture the third national lockdown beginning 5 January 2021. Dark grey regions indicate national lockdowns, light grey indicate the period where London and much of the South East and East of England were placed in Tier 4 restrictions.

Discussion

Using data collected through community reporting of symptoms and tests via the COVID Symptom Study app, we investigated whether the appearance of the variant B.1.1.7, first detected in a sample from England in September 2020, was related to differences in symptom reporting, disease duration, hospitalisation, asymptomatic infection, risks of reinfection, and transmissibility for users reporting a positive test result between 28 September and 27 December 2020.

We did not find associations between the proportion of B.1.1.7 in circulation and disease severity, either measured by the number of different reported symptoms over the 4-week window around each positive test, hospitalisations, any of the different symptoms, or the proportion of individuals with long symptom duration when correcting for variations in demographic characteristics (age, sex) and seasonal variables (temperature, humidity). The proportion of individuals with duration of symptoms ≥ 28 days without a break of more than seven days did not change in association with the presence of variant B.1.1.7. Likewise the proportion of users with asymptomatic disease did not significantly change as B.1.1.7 increased in prevalence.

A recent study reported that individuals infected with B.1.1.7 were more likely to report a cough, sore throat, fatigue, myalgia and fever in the seven days preceding the test, and less likely to report a loss of taste or smell.¹³ It is not clear if this report adjusted for age, sex, and environmental factors. If we do not correct for these factors we find some significant changes in symptom reporting but in our view these are not likely to be due to B.1.1.7 (Supplementary Figure 4). The periods considered also differed; we considered symptoms reported both two weeks before and after the positive test result. Further opportunity to study symptoms with B.1.1.7 in different contexts are required to be definitive.

We observed, based on 249 potential cases, a very low rate of possible reinfection of 0.7% (95% CI 0.6- 0.8). This rate is consistent with another study of 6614 healthcare workers that had previously tested positive for Covid-19, finding 44 possible reinfections (0.66%).¹⁴ Our reinfection rate did not vary consistently across regions or time, which would be consistent with the hypothesis that reinfection is no more likely in the context of B.1.1.7. This may mean that if adequate immunity is built over the first infection it may be sufficient to protect against reinfection in the presence of B.1.1.7. Ultimately this is a positive sign that the immunity built through vaccination against the old variants could also be useful against B.1.1.7. This is in line with initial reports regarding the efficacy of vaccines designed for early strains against this newer variant.^{15,16}

We found an increase in the reproduction number $R(t)$ in association with the B.1.1.7 variant: we found a multiplicative increase in $R(t)$ of ~ 1.35 (95% 1.02-1.69), compatible with estimates from Volz et al.^{3,4}, of 1.4-1.8, and Davies et al. who estimated a transmissibility increase of 1.56 (95% CI 1.50-1.74).^{3,4} These increases in transmissivity have worrying implications for the ability of lockdown measures to control B.1.1.7, given $R(t)$ was estimated to be 0.7-0.9 during the first national lockdown in England.¹⁷ Despite this, we found $R(t)$ to be ~ 0.8 in the three regions in England with at least 80% of B.1.1.7, with very clear response to lockdown measures. This could indicate that the true increase in transmissivity is at the lower end of the available estimates, or that the increase in transmissivity estimated outside of lockdown cannot be extrapolated to lockdown, perhaps due to B.1.1.7 responding differently to lockdown measures than the old variants.

Strengths

The large, longitudinal nature of the CSS data, with good coverage of the UK population, provides a unique opportunity to study potential changes in symptomatology, symptom severity, and disease duration. The ability to match tests and symptom reports over long periods further allows us to measure possible reinfection rates. Our data also offers the ability to provide a valuable complementary measure to existing measurements of the increased transmissibility of B.1.1.7: we were able to use real-time, representative incidence estimates to measure $R(t)$, whilst other studies have relied on deaths and hospitalisations, which are lagged, or community case numbers which do not reflect true infection numbers.

Limitations

Our study is limited by reliance on self-report of symptoms and test results, although previous publications from our group show that our figures triangulate well with other study designs¹¹. Despite the ability of the app users to correct any wrong input of their test results, errors still may be made. We make the assumption that testing positive for SARS CoV2 after an interval of 90 days with at least seven days of freedom from symptoms in the interval is consistent with reinfection. Repeated positive testing has been reported shortly after hospital discharge¹⁸ and showed that PCR positivity could be detected up to 28 days post symptom resolution. While the chosen cut-off of 90 days between two positive tests is unlikely to be due to prolonged PCR positivity, this cannot be ruled out, but would only affect a small number of cases. Viral sequencing of the two infections would ideally be required to confirm reinfection. Despite correcting for changes in temperature and humidity, a possible limitation in the study is that comparisons in symptoms are made across time, and seasonal effects (e.g. on symptoms) may not have been fully taken into account¹⁹. As we lack information on the disease strain of individual positive infections reported through the app, the study is associative in nature; and we cannot account for the effects of other potentially circulating variants.

Conclusions

We examined the effect of SARS-CoV-2 variant B.1.1.7 on the symptoms, disease course, rates of reinfection, and transmissibility in the UK. We found no change in symptoms and no increase in overall disease severity. We found a low rate of reinfection (0.7%) and no evidence of increased rates associated with B.1.1.7. We found an increase in $R(t)$ of ~ 1.38 (95% CI 1.06-1.71), but evidence that lockdown measures are effective even in regions with very high (>80%) proportions of B.1.1.7.

Ethics

Ethics has been approved by KCL Ethics Committee REMAS ID 18210, review reference LRS-19/20-18210 and all participants provided consent.

Data sharing

Data collected in the COVID Symptom Study smartphone application are being shared with other health researchers through the UK National Health Service-funded Health Data Research UK (HDRUK) and Secure Anonymised Information Linkage consortium, housed in the UK Secure Research Platform (Swansea, UK). Anonymised data are available to be shared with researchers according to their protocols in the public interest (<https://web.www.healthdatagateway.org/dataset/fddcb382-3051-4394-8436-b92295f14259>). US investigators are encouraged to coordinate data requests through the Coronavirus Pandemic Epidemiology Consortium (<https://www.monganinstitute.org/cope-consortium>).

Acknowledgements

ZOE Global provided in kind support for all aspects of building, running and supporting the app and service to all users worldwide. COG-UK is supported by funding from the Medical Research

Council (MRC) part of UK Research & Innovation (UKRI), the National Institute of Health Research (NIHR) and Genome Research Limited, operating as the Wellcome Sanger Institute. Support for this study was provided by the NIHR-funded Biomedical Research Centre based at GSTT NHS Foundation Trust. Investigators also received support from the Wellcome Trust, the MRC/BHF, Alzheimer's Society, EU, NIHR, CDRF, and the NIHR-funded BioResource, Clinical Research Facility and BRC based at GSTT NHS Foundation Trust in partnership with KCL, the UK Research and Innovation London Medical Imaging & Artificial Intelligence Centre for Value Based Healthcare, the Wellcome Flagship Programme (WT213038/Z/18/Z), the Chronic Disease Research Foundation, and DHSC. ATC was supported in this work through a Stuart and Suzanne Steele MGH Research Scholar Award. The Massachusetts Consortium on Pathogen Readiness (MassCPR) and Mark and Lisa Schwartz supported MGH investigators (DAD, LHN, ATC).

Declaration of interests

AM, LP, SS, JCP, CH, JW are employees of Zoe Global Ltd. TDS is a consultant to Zoe Global Ltd. DAD and ATC previously served as investigators on a clinical trial of diet and lifestyle using a separate smartphone application that was supported by Zoe Global.

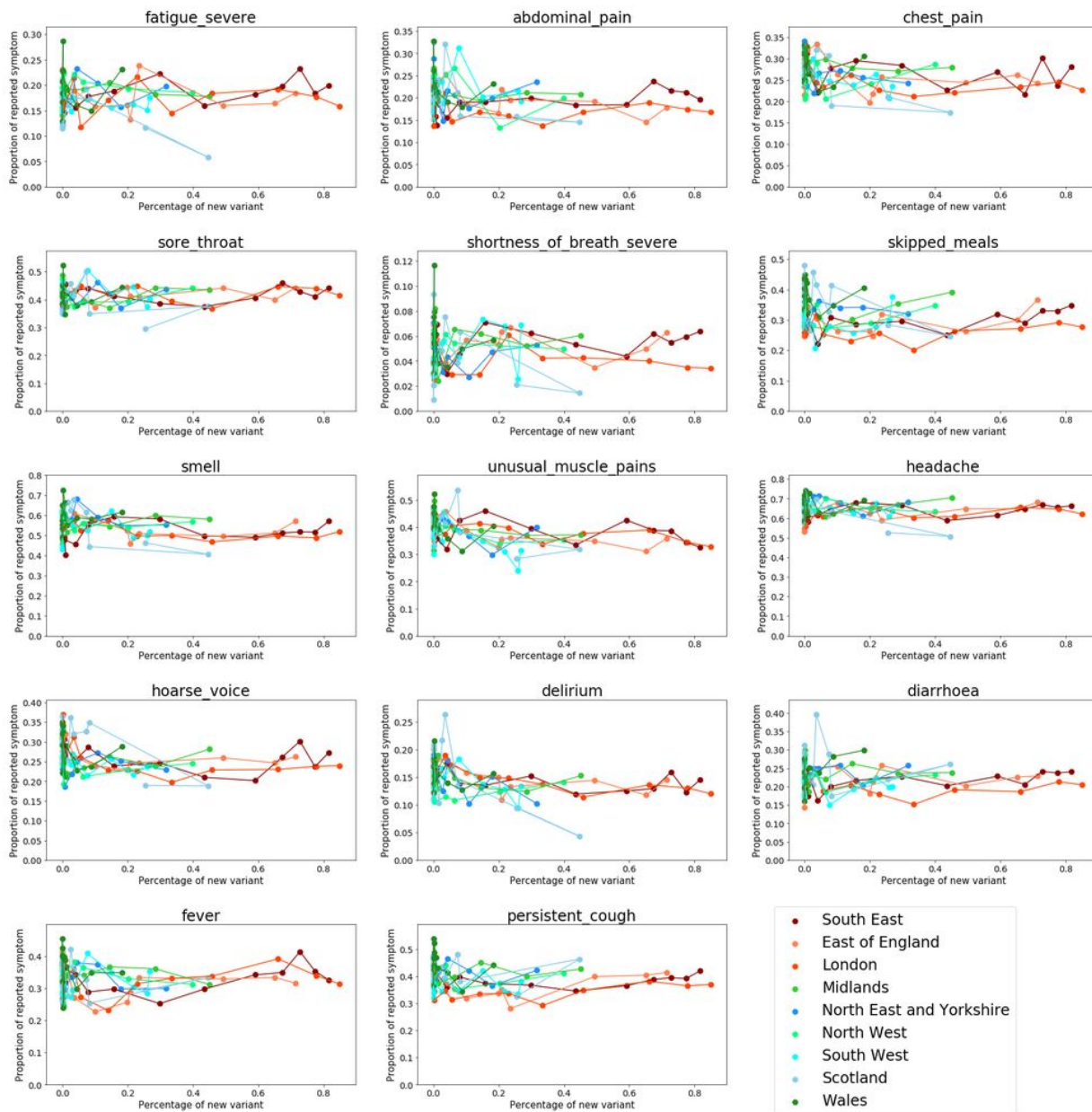
References

- 1 Public Health England. Investigation of novel SARS-COV-2 variant Variant of Concern 202012/01 Technical briefing 1. 2020.
- 2 Public Health England. Investigation of Novel SARS-COV-2 Variant Variant of Concern 202012/01 Technical Briefing 2. 2020.
- 3 Davies NG, Barnard RC, Jarvis CI, *et al.* Estimated transmissibility and severity of novel SARS-CoV-2 Variant of Concern 202012/01 in England. *medRxiv* 2020. <https://www.medrxiv.org/content/10.1101/2020.12.24.20248822v1.full-text>.
- 4 Volz E, Mishra S, Chand M, *et al.* Transmission of SARS-CoV-2 Lineage B.1.1.7 in England: Insights from linking epidemiological and genetic data. DOI:10.1101/2020.12.30.20249034.
- 5 Peter Horby, Catherine Huntley, Nick Davies, John Edmunds, Neil Ferguson, Graham Medley, Andrew Hayward, Muge Cevik, Calum Semple. NERVTAG note on B.1.1.7 severity. NERVTAG, 2021.
- 6 Drew DA, Nguyen LH, Steves CJ, *et al.* Rapid implementation of mobile technology for real-time epidemiology of COVID-19. *Science* 2020; **368**: 1362–7.

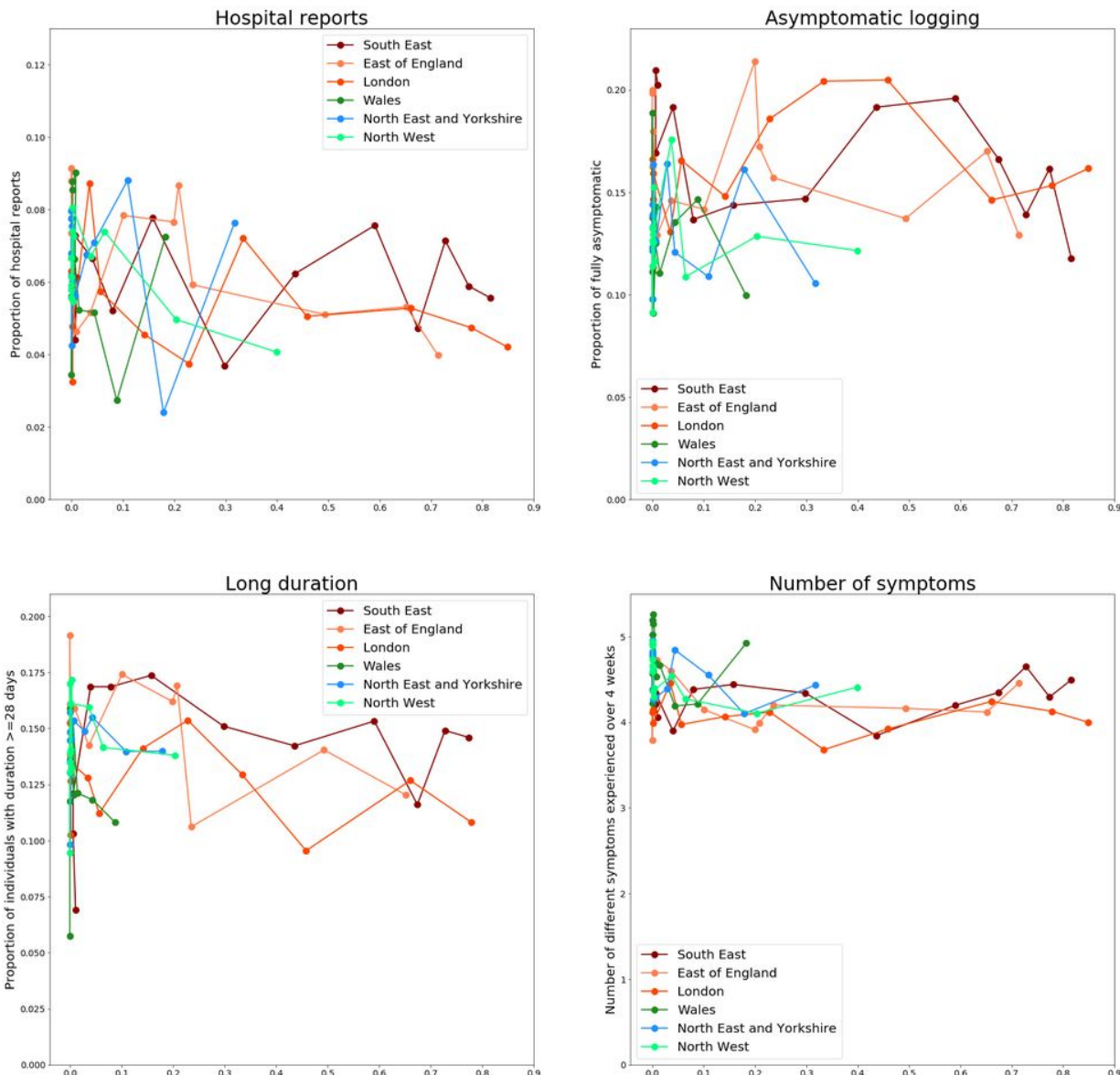
- 7 COVID-19 Genomics UK (COG-UK) consortiumcontact@cogconsortium.uk. An integrated national scale SARS-CoV-2 genomic surveillance network. *Lancet Microbe* 2020; **1**: e99–100.
- 8 Public Health England. Investigation of Novel SARS-COV-2 Variant Variant of Concern 202012/01 Technical Briefing 3. 2021.
- 9 NASA. NASA POWER Climate Data. <https://power.larc.nasa.gov/> (accessed Jan 25, 2021).
- 10 Sudre CH, Murray B, Varsavsky T, *et al.* Attributes and predictors of Long-COVID: analysis of COVID cases and their symptoms collected by the Covid Symptoms Study App. DOI:10.1101/2020.10.19.20214494.
- 11 Varsavsky T, Graham MS, Canas LS, *et al.* Detecting COVID-19 infection hotspots in England using large-scale self-reported data from a mobile application: a prospective, observational study. *The Lancet Public Health* 2020; published online Dec 3. DOI:10.1016/S2468-2667(20)30269-3.
- 12 Public Health England. UK Covid-19 Dashboard. <https://coronavirus.data.gov.uk/> (accessed Jan 19, 2021).
- 13 Office for National Statistics. Coronavirus (COVID-19) Infection Survey: characteristics of people testing positive for COVID-19 in England. 2021 <https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/conditionsanddiseases/articles/coronaviruscovid19infectionsinthecommunityinengland/characteristicsofpeopletestingpositiveforcovid19inengland27january2021>.
- 14 Hall V, Foulkes S, Charlett A, *et al.* Do antibody positive healthcare workers have lower SARS-CoV-2 infection rates than antibody negative healthcare workers? Large multi-centre prospective cohort study (the SIREN study), England: June to November 2020. bioRxiv. 2021; published online Jan 15. DOI:10.1101/2021.01.13.21249642.
- 15 Xie X, Zou J, Fontes-Garfias CR, *et al.* Neutralization of N501Y mutant SARS-CoV-2 by BNT162b2 vaccine-elicited sera. Cold Spring Harbor Laboratory. 2021; : 2021.01.07.425740.
- 16 Wu K, Werner AP, Moliva JI, *et al.* mRNA-1273 vaccine induces neutralizing antibodies against spike mutants from global SARS-CoV-2 variants. DOI:10.1101/2021.01.25.427948.
- 17 UK Government R estimates. <https://www.gov.uk/guidance/the-r-number-in-the-uk> (accessed Jan 21, 2020).
- 18 Zheng J, Zhou R, Chen F, *et al.* Incidence, clinical course and risk factor for recurrent PCR positivity in discharged COVID-19 patients in Guangzhou, China: A prospective cohort study. *PLOS Neglected Tropical Diseases*. 2020; **14**: e0008648.
- 19 Kifer D, Bugada D, Villar-Garcia J, *et al.* Effects of environmental factors on severity and mortality of COVID-19. *MedRxiv* 2020. <https://www.medrxiv.org/content/10.1101/2020.07.11.20147157v3.full-text>.

Supplementary material

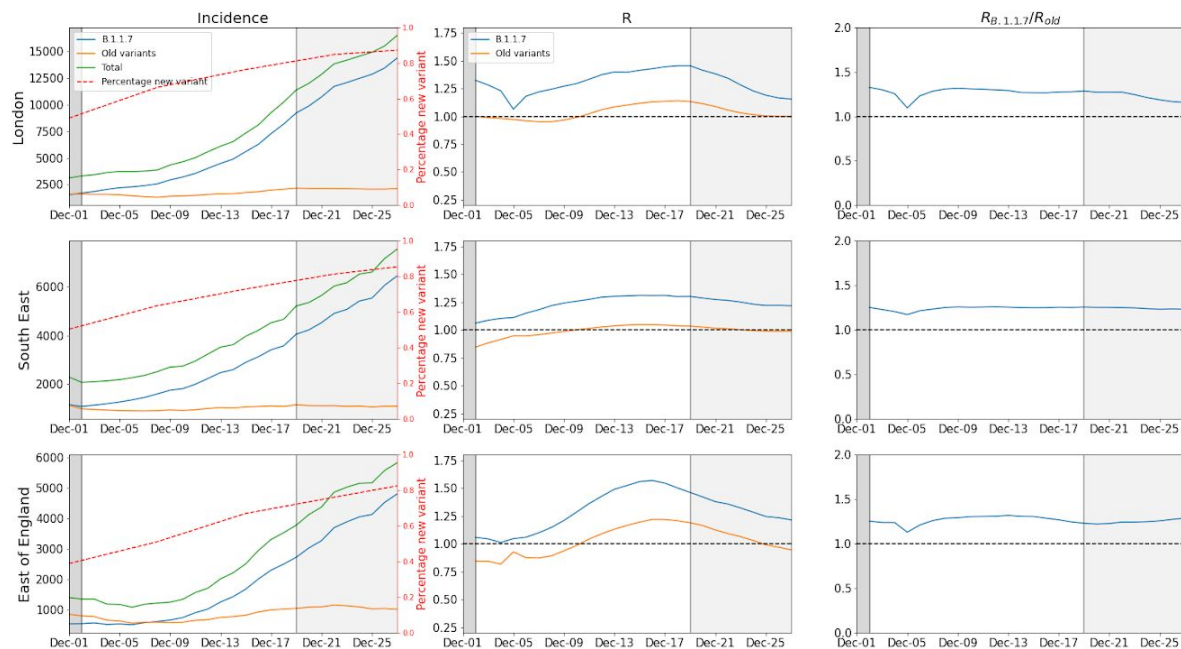
Supplementary tables and figures



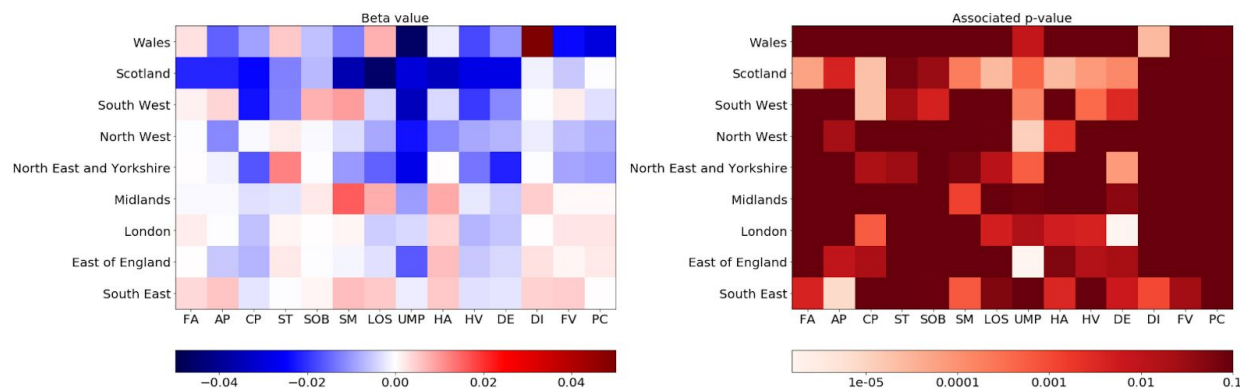
Supplementary Figure 1. Regional plots of the frequency of reporting of symptoms over time for each reported symptom, against the proportion of B.1.1.7.. Drop in fever reporting in early November was caused by a change in the question wording; this wording was subsequently reverted a week later.



Supplementary Figure 2. Regional plot of hospitalisation report, proportion of asymptomatic, report of long duration and number of experienced symptoms around test against proportion of B.1.1.7. For the study of long duration, tests are only considered up to 21 December counting reports up to 18 January 2021 to limit right censoring. Only symptomatic individuals for which duration can be ascertained are included.



Supplementary Figure 3. Incidence and $R(t)$ for the old and new variants, along with the ratio between these R values, for the three regions in England with the largest proportion of B.1.1.7, using SGTF data. Dark grey regions indicate national lockdowns, light grey shaded the period where London and much of the South East and East of England were placed in Tier 4 restrictions.



Supplementary Figure 4: Colour plot of beta values and associated p-values for each region and symptoms when investigating association between symptom report (in a 4 week window around the test) and proportion of variant B.1.1.7 and without any correction for personal characteristic or seasonal feature. Note that the p-values are capped at 0.1. Beta-values are presented for an increase of 0.1 in the proportion of variant B.1.1.7.

Key: FA - fatigue, AP - abdominal pain, CP - chest pain, ST - sore throat, SOB - shortness of breath, SM - skipped meals, LOS - loss of smell, UMP - unusual muscle pains, HA - headache, HV - hoarse voice, DE - delirium, DI - diarrhoea, FV - fever, PC - persistent cough

	Proportion of fully asymptomatic	Number of symptoms reported over 4 weeks around test	Proportion of hospital reports	Proportion of individuals with symptom duration \geq 28days
South East	0.001 [-0.015;0.017] ; 0.901	-0.021 [-0.163;0.121] ; 0.733	-0.002 [-0.011;0.007] ; 0.624	-0.003 [-0.009;0.004] ; 0.37
East of England	0.002 [-0.008;0.012] ; 0.588	-0.012 [-0.153;0.13] ; 0.851	-0.002 [-0.01;0.006] ; 0.52	-0.002 [-0.015;0.01] ; 0.689
London	-0.005 [-0.014;0.005] ; 0.298	0.031 [-0.055;0.116] ; 0.423	-0.002 [-0.007;0.003] ; 0.298	-0.002 [-0.013;0.009] ; 0.682
Midlands	-0.016 [-0.028;-0.004] ; 0.014	0.02 [-0.133;0.173] ; 0.766	-0.002 [-0.007;0.003] ; 0.328	0.002 [-0.01;0.015] ; 0.671
North East and Yorkshire	-0.011 [-0.046;0.023] ; 0.462	-0.086 [-0.444;0.272] ; 0.586	-0.011 [-0.04;0.019] ; 0.426	0.015 [-0.022;0.052] ; 0.349
North West	-0.005 [-0.023;0.013] ; 0.512	-0.053 [-0.218;0.111] ; 0.468	-0.009 [-0.015;-0.004] ; 0.005	0 [-0.031;0.031] ; 0.98
South West	0.015 [-0.011;0.04] ; 0.217	-0.261 [-0.437;-0.085] ; 0.01	-0.001 [-0.02;0.018] ; 0.902	-0.048 [-0.091;-0.004] ; 0.036
Scotland	0.022 [-0.013;0.058] ; 0.177	-0.4 [-0.711;-0.088] ; 0.019	-0.018 [-0.037;0.002] ; 0.073	-0.012 [-0.027;0.003] ; 0.107
Wales	-0.002 [-0.05;0.047] ; 0.943	-0.041 [-0.683;0.601] ; 0.884	-0.008 [-0.045;0.028] ; 0.602	-0.053 [-0.141;0.035] ; 0.192

Supplementary Table 1: Beta coefficient of the variant proportion when evaluating association with number of reported symptoms, asymptomatic rate, proportion of hospital report and proportion of individuals with duration >28 days (among symptomatic) across the different regions when correcting for age, sex, temperature and humidity. All values are presented for an increase in 0.1 in the proportion of variant B.1.1.7. All results are presented in the form mean [CI]; p-value

Region	Correlation Variant/Reinfection	Correlation New cases/Reinfection	p-value
South East	0.55	0.69	<0.001
East of England	0.51	0.56	<0.001
London	0.46	0.62	<0.001
Midlands	0.28	0.75	<0.001
North East and Yorkshire	-0.02	0.30	<0.001
North West	0.06	0.43	<0.001
South West	-0.35	0.05	<0.001
Scotland	0.59	-0.15	<0.001
Wales	0.07	0.26	<0.001

Supplementary Table 2: Comparison of regional correlation over time between proportion of B.1.1.7 and number of possible reinfections and between new reported cases and number of possible reinfections. Medians over 100 bootstrapped samples are calculated for each and compared using a Mann-Whitney U test.

COG-UK authorship list

Funding acquisition, Leadership and supervision, Metadata curation, Project administration, Samples and logistics, Sequencing and analysis, Software and analysis tools, and Visualisation:
Samuel C Robson ¹³.

Funding acquisition, Leadership and supervision, Metadata curation, Project administration, Samples and logistics, Sequencing and analysis, and Software and analysis tools:
Nicholas J Loman ⁴¹ and Thomas R Connor ^{10, 69}.

Leadership and supervision, Metadata curation, Project administration, Samples and logistics, Sequencing and analysis, Software and analysis tools, and Visualisation:
Tanya Golubchik ⁵.

Funding acquisition, Metadata curation, Samples and logistics, Sequencing and analysis, Software and analysis tools, and Visualisation:
Rocio T Martinez Nunez ⁴².

Funding acquisition, Leadership and supervision, Metadata curation, Project administration, and Samples and logistics:
Catherine Ludden ⁸⁸.

Funding acquisition, Leadership and supervision, Metadata curation, Samples and logistics, and Sequencing and analysis:
Sally Corden ⁶⁹.

Funding acquisition, Leadership and supervision, Project administration, Samples and logistics, and Sequencing and analysis:
Ian Johnston ⁹⁹ and David Bonsall ⁵.

Funding acquisition, Leadership and supervision, Sequencing and analysis, Software and analysis tools, and Visualisation:
Colin P Smith ⁸⁷ and Ali R Awan ²⁸.

Funding acquisition, Samples and logistics, Sequencing and analysis, Software and analysis tools, and Visualisation:
Giselda Bucca ⁸⁷.

Leadership and supervision, Metadata curation, Project administration, Samples and logistics, and Sequencing and analysis:
M. Estee Torok ^{22, 101}.

Leadership and supervision, Metadata curation, Project administration, Samples and logistics, and Visualisation:
Kordo Saeed ^{81, 110} and Jacqui A Prieto ^{83, 109}.

Leadership and supervision, Metadata curation, Project administration, Sequencing and analysis, and Software and analysis tools:

David K Jackson ⁹⁹.

Metadata curation, Project administration, Samples and logistics, Sequencing and analysis, and Software and analysis tools:

William L Hamilton ²².

Metadata curation, Project administration, Samples and logistics, Sequencing and analysis, and Visualisation:

Luke B Snell ¹¹.

Funding acquisition, Leadership and supervision, Metadata curation, and Samples and logistics:

Catherine Moore ⁶⁹.

Funding acquisition, Leadership and supervision, Project administration, and Samples and logistics:

Ewan M Harrison ^{99, 88}.

Leadership and supervision, Metadata curation, Project administration, and Samples and logistics:

Sonia Goncalves ⁹⁹.

Leadership and supervision, Metadata curation, Samples and logistics, and Sequencing and analysis:

Ian G Goodfellow ²⁴, Derek J Fairley ^{3, 72}, Matthew W Loose ¹⁸ and Joanne Watkins ⁶⁹.

Leadership and supervision, Metadata curation, Samples and logistics, and Software and analysis tools:

Rich Livett ⁹⁹.

Leadership and supervision, Metadata curation, Samples and logistics, and Visualisation:

Samuel Moses ^{25, 106}.

Leadership and supervision, Metadata curation, Sequencing and analysis, and Software and analysis tools:

Roberto Amato ⁹⁹, Sam Nicholls ⁴¹ and Matthew Bull ⁶⁹.

Leadership and supervision, Project administration, Samples and logistics, and Sequencing and analysis:

Darren L Smith ^{37, 58, 105}.

Leadership and supervision, Sequencing and analysis, Software and analysis tools, and Visualisation:

Jeff Barrett ⁹⁹, David M Aanensen ^{14, 114}.

Metadata curation, Project administration, Samples and logistics, and Sequencing and analysis:

Martin D Curran ⁶⁵, Surendra Parmar ⁶⁵, Dinesh Aggarwal ^{95, 99, 64} and James G Shepherd ⁴⁸.

Metadata curation, Project administration, Sequencing and analysis, and Software and analysis tools:

Matthew D Parker ⁹³.

Metadata curation, Samples and logistics, Sequencing and analysis, and Visualisation:

Sharon Glaysher ⁶¹.

Metadata curation, Sequencing and analysis, Software and analysis tools, and Visualisation:

Matthew Bashton ^{37, 58}, Anthony P Underwood ^{14, 114}, Nicole Pacchiarini ⁶⁹ and Katie F Loveson ⁷⁷.

Project administration, Sequencing and analysis, Software and analysis tools, and Visualisation:

Alessandro M Carabelli ⁸⁸.

Funding acquisition, Leadership and supervision, and Metadata curation:

Kate E Templeton ^{53, 90}.

Funding acquisition, Leadership and supervision, and Project administration:

Cordelia F Langford ⁹⁹, John Sillitoe ⁹⁹, Thushan I de Silva ⁹³ and Dennis Wang ⁹³.

Funding acquisition, Leadership and supervision, and Sequencing and analysis:

Dominic Kwiatkowski ^{99, 107}, Andrew Rambaut ⁹⁰, Justin O'Grady ^{70, 89} and Simon Cottrell ⁶⁹.

Leadership and supervision, Metadata curation, and Sequencing and analysis:

Matthew T.G. Holden ⁶⁸ and Emma C Thomson ⁴⁸.

Leadership and supervision, Project administration, and Samples and logistics:

Husam Osman ^{64, 36}, Monique Andersson ⁵⁹, Anoop J Chauhan ⁶¹ and Mohammed O Hassan-Ibrahim ⁶.

Leadership and supervision, Project administration, and Sequencing and analysis:

Mara Lawniczak ⁹⁹.

Leadership and supervision, Samples and logistics, and Sequencing and analysis:

Ravi Kumar Gupta ^{88, 113}, Alex Alderton ⁹⁹, Meera Chand ⁶⁶, Chrystala Constantinidou ⁹⁴, Meera Unnikrishnan ⁹⁴, Alistair C Darby ⁹², Julian A Hiscox ⁹² and Steve Paterson ⁹².

Leadership and supervision, Sequencing and analysis, and Software and analysis tools:

Inigo Martincorena ⁹⁹, David L Robertson ⁴⁸, Erik M Volz ³⁹, Andrew J Page ⁷⁰ and Oliver G Pybus ²³.

Leadership and supervision, Sequencing and analysis, and Visualisation:

Andrew R Bassett ⁹⁹.

Metadata curation, Project administration, and Samples and logistics:

Cristina V Ariani ⁹⁹, Michael H Spencer Chapman ^{99, 88}, Kathy K Li ⁴⁸, Rajiv N Shah ⁴⁸, Natasha G Jesudason ⁴⁸ and Yusri Taha ⁵⁰.

Metadata curation, Project administration, and Sequencing and analysis:

Martin P McHugh ⁵³ and Rebecca Dewar ⁵³.

Metadata curation, Samples and logistics, and Sequencing and analysis:

Aminu S Jahun²⁴, Claire McMurray⁴¹, Sarojini Pandey⁸⁴, James P McKenna³, Andrew Nelson^{58, 105}, Gregory R Young^{37, 58}, Clare M McCann^{58, 105} and Scott Elliott⁶¹.

Metadata curation, Samples and logistics, and Visualisation:

Hannah Lowe²⁵.

Metadata curation, Sequencing and analysis, and Software and analysis tools:

Ben Temperton⁹¹, Sunando Roy⁸², Anna Price¹⁰, Sara Rey⁶⁹ and Matthew Wyles⁹³.

Metadata curation, Sequencing and analysis, and Visualisation:

Stefan Rooke⁹⁰ and Sharif Shaaban⁶⁸.

Project administration, Samples and logistics, Sequencing and analysis:

Mariateresa de Cesare⁹⁸.

Project administration, Samples and logistics, and Software and analysis tools:

Laura Letchford⁹⁹.

Project administration, Samples and logistics, and Visualisation:

Siona Silveira⁸¹, Emanuela Pelosi⁸¹ and Eleri Wilson-Davies⁸¹.

Samples and logistics, Sequencing and analysis, and Software and analysis tools:

Myra Hosmillo²⁴.

Sequencing and analysis, Software and analysis tools, and Visualisation:

Áine O'Toole⁹⁰, Andrew R Hesketh⁸⁷, Richard Stark⁹⁴, Louis du Plessis²³, Chris Ruis⁸⁸, Helen Adams⁴ and Yann Bourgeois⁷⁶.

Funding acquisition, and Leadership and supervision:

Stephen L Michell⁹¹, Dimitris Grammatopoulos^{84, 112}, Jonathan Edgeworth¹², Judith Breuer^{30, 82}, John A Todd⁹⁸ and Christophe Fraser⁵.

Funding acquisition, and Project administration:

David Buck⁹⁸ and Michaela John⁹.

Leadership and supervision, and Metadata curation:

Gemma L Kay⁷⁰.

Leadership and supervision, and Project administration:

Steve Palmer⁹⁹, Sharon J Peacock^{88, 64} and David Heyburn⁶⁹.

Leadership and supervision, and Samples and logistics:

Danni Weldon⁹⁹, Esther Robinson^{64, 36}, Alan McNally^{41, 86}, Peter Muir⁶⁴, Ian B Vipond⁶⁴, John BoYes²⁹, Venkat Sivaprakasam⁴⁶, Tranprit Salluja⁷⁵, Samir Dervisevic⁵⁴ and Emma J Meader⁵⁴.

Leadership and supervision, and Sequencing and analysis:

Naomi R Park⁹⁹, Karen Oliver⁹⁹, Aaron R Jeffries⁹¹, Sascha Ott⁹⁴, Ana da Silva Filipe⁴⁸, David A Simpson⁷² and Chris Williams⁶⁹.

Leadership and supervision, and Visualisation:

Jane AH Masoli^{73, 91}.

Metadata curation, and Samples and logistics:

Bridget A Knight^{73, 91}, Christopher R Jones^{73, 91}, Cherian Koshy¹, Amy Ash¹, Anna Casey⁷¹, Andrew Bosworth^{64, 36}, Liz Ratcliffe⁷¹, Li Xu-McCrae³⁶, Hannah M Pymont⁶⁴, Stephanie Hutchings⁶⁴, Lisa Berry⁸⁴, Katie Jones⁸⁴, Fenella Halstead⁴⁶, Thomas Davis²¹, Christopher Holmes¹⁶, Miren Iturriza-Gomara⁹², Anita O Lucaci⁹², Paul Anthony Randell^{38, 104}, Alison Cox^{38, 104}, Pinglawathee Madona^{38, 104}, Kathryn Ann Harris³⁰, Julianne Rose Brown³⁰, Tabitha W Mahungu⁷⁴, Dianne Irish-Tavares⁷⁴, Tanzina Haque⁷⁴, Jennifer Hart⁷⁴, Eric Witele⁷⁴, Melisa Louise Fenton⁷⁵, Steven Liggett⁷⁹, Clive Graham⁵⁶, Emma Swindells⁵⁷, Jennifer Collins⁵⁰, Gary Eltringham⁵⁰, Sharon Campbell¹⁷, Patrick C McClure⁹⁷, Gemma Clark¹⁵, Tim J Sloan⁶⁰, Carl Jones¹⁵ and Jessica Lynch^{2, 111}.

Metadata curation, and Sequencing and analysis:

Ben Warne⁸, Steven Leonard⁹⁹, Jillian Durham⁹⁹, Thomas Williams⁹⁰, Sam T Haldenby⁹², Nathaniel Storey³⁰, Nabil-Fareed Alikhan⁷⁰, Nadine Holmes¹⁸, Christopher Moore¹⁸, Matthew Carlile¹⁸, Malorie Perry⁶⁹, Noel Craine⁶⁹, Ronan A Lyons⁸⁰, Angela H Beckett¹³, Salman Goudarzi⁷⁷, Christopher Fearn⁷⁷, Kate Cook⁷⁷, Hannah Dent⁷⁷ and Hannah Paul⁷⁷.

Metadata curation, and Software and analysis tools:

Robert Davies⁹⁹.

Project administration, and Samples and logistics:

Beth Blane⁸⁸, Sophia T Girgis⁸⁸, Mathew A Beale⁹⁹, Katherine L Bellis^{99, 88}, Matthew J Dorman⁹⁹, Eleanor Drury⁹⁹, Leanne Kane⁹⁹, Sally Kay⁹⁹, Samantha McGuigan⁹⁹, Rachel Nelson⁹⁹, Liam Prestwood⁹⁹, Shavanthi Rajatileka⁹⁹, Rahul Batra¹², Rachel J Williams⁸², Mark Kristiansen⁸², Angie Green⁹⁸, Anita Justice⁵⁹, Adhyana I.K Mahanama^{81, 102} and Buddhini Samaraweera^{81, 102}.

Project administration, and Sequencing and analysis:

Nazreen F Hadjirin⁸⁸ and Joshua Quick⁴¹.

Project administration, and Software and analysis tools:

Radoslaw Poplawski⁴¹.

Samples and logistics, and Sequencing and analysis:

Leanne M Kermack⁸⁸, Nicola Reynolds⁷, Grant Hall²⁴, Yasmin Chaudhry²⁴, Malte L Pinckert²⁴, Iliana Georgana²⁴, Robin J Moll⁹⁹, Alicia Thornton⁶⁶, Richard Myers⁶⁶, Joanne Stockton⁴¹, Charlotte A Williams⁸², Wen C Yew⁵⁸, Alexander J Trotter⁷⁰, Amy Trebes⁹⁸, George MacIntyre-Cockett⁹⁸, Alec Birchley⁶⁹, Alexander Adams⁶⁹, Amy Plimmer⁶⁹, Bree Gatica-Wilcox⁶⁹, Caoimhe McKerr⁶⁹, Ember Hilvers⁶⁹, Hannah Jones⁶⁹, Hibo Asad⁶⁹, Jason Coombes⁶⁹, Johnathan M Evans⁶⁹, Laia Fina⁶⁹, Lauren Gilbert⁶⁹, Lee Graham⁶⁹, Michelle Cronin⁶⁹, Sara Kumziene-SummerhaYes⁶⁹, Sarah Taylor⁶⁹, Sophie Jones⁶⁹, Danielle C Groves⁹³, Peijun Zhang⁹³, Marta Gallis⁹³ and Stavroula F Louka⁹³.

Samples and logistics, and Software and analysis tools:

Igor Starinskij ⁴⁸.

Sequencing and analysis, and Software and analysis tools:

Chris J Illingworth ⁴⁷, Chris Jackson ⁴⁷, Marina Gourtovaia ⁹⁹, Gerry Tonkin-Hill ⁹⁹, Kevin Lewis ⁹⁹, Jaime M Tovar-Corona ⁹⁹, Keith James ⁹⁹, Laura Baxter ⁹⁴, Mohammad T. Alam ⁹⁴, Richard J Orton ⁴⁸, Joseph Hughes ⁴⁸, Sreenu Vattipally ⁴⁸, Manon Ragonnet-Cronin ³⁹, Fabricia F. Nascimento ³⁹, David Jorgensen ³⁹, Olivia Boyd ³⁹, Lily Geidelberg ³⁹, Alex E Zarebski ²³, Jayna Raghvani ²³, Moritz UG Kraemer ²³, Joel Southgate ^{10, 69}, Benjamin B Lindsey ⁹³ and Timothy M Freeman ⁹³.

Software and analysis tools, and Visualisation:

Jon-Paul Keatley ⁹⁹, Joshua B Singer ⁴⁸, Leonardo de Oliveira Martins ⁷⁰, Corin A Yeats ¹⁴, Khalil Abudahab ^{14, 114}, Ben EW Taylor ^{14, 114} and Mirko Menegazzo ¹⁴.

Leadership and supervision:

John Danesh ⁹⁹, Wendy Hogsdon ⁴⁶, Sahar Eldirdiri ²¹, Anita Kenyon ²¹, Jenifer Mason ⁴³, Trevor I Robinson ⁴³, Alison Holmes ^{38, 103}, James Price ^{38, 103}, John A Hartley ⁸², Tanya Curran ³, Alison E Mather ⁷⁰, Giri Shankar ⁶⁹, Rachel Jones ⁶⁹, Robin Howe ⁶⁹ and Sian Morgan ⁹.

Metadata curation:

Elizabeth Wastenge ⁵³, Michael R Chapman ^{34, 88, 99}, Siddharth Mookerjee ^{38, 103}, Rachael Stanley ⁵⁴, Wendy Smith ¹⁵, Timothy Peto ⁵⁹, David Eyre ⁵⁹, Derrick Crook ⁵⁹, Gabrielle Vernet ³³, Christine Kitchen ¹⁰, Huw Gulliver ¹⁰, Ian Merrick ¹⁰, Martyn Guest ¹⁰, Robert Munn ¹⁰, Declan T Bradley ^{63, 72} and Tim Wyatt ⁶³.

Project administration:

Charlotte Beaver ⁹⁹, Luke Foulser ⁹⁹, Sophie Palmer ⁸⁸, Carol M Churcher ⁸⁸, Ellena Brooks ⁸⁸, Kim S Smith ⁸⁸, Katerina Galai ⁸⁸, Georgina M McManus ⁸⁸, Frances Bolt ^{38, 103}, Francesc Coll ¹⁹, Lizzie Meadows ⁷⁰, Stephen W Attwood ²³, Alisha Davies ⁶⁹, Elen De Lacy ⁶⁹, Fatima Downing ⁶⁹, Sue Edwards ⁶⁹, Garry P Scarlett ⁷⁶, Sarah Jeremiah ⁸³ and Nikki Smith ⁹³.

Samples and logistics:

Danielle Leek ⁸⁸, Sushmita Sridhar ^{88, 99}, Sally Forrest ⁸⁸, Claire Cormie ⁸⁸, Harmeet K Gill ⁸⁸, Joana Dias ⁸⁸, Ellen E Higginson ⁸⁸, Mailis Maes ⁸⁸, Jamie Young ⁸⁸, Michelle Wantoch ⁷, Sanger Covid Team (www.sanger.ac.uk/covid-team) ⁹⁹, Dorota Jamrozny ⁹⁹, Stephanie Lo ⁹⁹, Minal Patel ⁹⁹, Verity Hill ⁹⁰, Claire M Bewshea ⁹¹, Sian Ellard ^{73, 91}, Cressida Auckland ⁷³, Ian Harrison ⁶⁶, Chloe Bishop ⁶⁶, Vicki Chalker ⁶⁶, Alex Richter ⁸⁵, Andrew Beggs ⁸⁵, Angus Best ⁸⁶, Benita Percival ⁸⁶, Jeremy Mirza ⁸⁶, Oliver Megram ⁸⁶, Megan Mayhew ⁸⁶, Liam Crawford ⁸⁶, Fiona Ashcroft ⁸⁶, Emma Moles-Garcia ⁸⁶, Nicola Cumley ⁸⁶, Richard Hopes ⁶⁴, Patawee Asamaphan ⁴⁸, Marc O Niebel ⁴⁸, Rory N Gunson ¹⁰⁰, Amanda Bradley ⁵², Alasdair Maclean ⁵², Guy Mollett ⁵², Rachel Blacow ⁵², Paul Bird ¹⁶, Thomas Helmer ¹⁶, Karlie Fallon ¹⁶, Julian Tang ¹⁶, Antony D Hale ⁴⁹, Louissa R Macfarlane-Smith ⁴⁹, Katherine L Harper ⁴⁹, Holli Carden ⁴⁹, Nicholas W Machin ^{45, 64}, Kathryn A Jackson ⁹², Shazaad S Y Ahmad ^{45, 64}, Ryan P George ⁴⁵, Lance Turtle ⁹², Elaine O'Toole ⁴³, Joanne Watts ⁴³, Cassie Breen ⁴³, Angela Cowell ⁴³, Adela Alcolea-Medina ^{32, 96}, Themoula Charalampous ^{12, 42}, Amita Patel ¹¹, Lisa J Levett ³⁵, Judith Heaney ³⁵, Aileen Rowan ³⁹, Graham P Taylor ³⁹, Divya Shah ³⁰, Laura Atkinson ³⁰, Jack CD Lee ³⁰, Adam P Westhorpe ⁸², Riaz Jannoo ⁸², Helen L Lowe ⁸², Angeliki Karamani ⁸², Leah Ensell ⁸², Wendy Chatterton ³⁵, Monika Pusok ³⁵, Ashok Dadrah ⁷⁵, Amanda Symmonds ⁷⁵, Graciela Sluga ⁴⁴, Zoltan Molnar ⁷², Paul Baker ⁷⁹, Stephen Bonner ⁷⁹, Sarah Essex ⁷⁹, Edward Barton ⁵⁶, Debra Padgett ⁵⁶, Garren Scott ⁵⁶, Jane Greenaway ⁵⁷, Brendan Al Payne ⁵⁰, Shirelle Burton-Fanning ⁵⁰, Sheila Waugh ⁵⁰, Veena Raviprakash ¹⁷, Nicola Sheriff ¹⁷, Victoria Blakey ¹⁷, Lesley-Anne Williams ¹⁷, Jonathan Moore ²⁷, Susanne Stonehouse ²⁷, Louise Smith ⁵⁵, Rose K Davidson

⁸⁹, Luke Bedford ²⁶, Lindsay Coupland ⁵⁴, Victoria Wright ¹⁸, Joseph G Chappell ⁹⁷, Theocharis Tsoleridis ⁹⁷, Jonathan Ball ⁹⁷, Manjinder Khakh ¹⁵, Vicki M Fleming ¹⁵, Michelle M Lister ¹⁵, Hannah C Howson-Wells ¹⁵, Louise Berry ¹⁵, Tim Boswell ¹⁵, Amelia Joseph ¹⁵, Iona Willingham ¹⁵, Nichola Duckworth ⁶⁰, Sarah Walsh ⁶⁰, Emma Wise ^{2, 111}, Nathan Moore ^{2, 111}, Matilde Mori ^{2, 108, 111}, Nick Cortes ^{2, 111}, Stephen Kidd ^{2, 111}, Rebecca Williams ³³, Laura Gifford ⁶⁹, Kelly Bicknell ⁶¹, Sarah Wyllie ⁶¹, Allyson Lloyd ⁶¹, Robert Impey ⁶¹, Cassandra S Malone ⁶, Benjamin J Cogger ⁶, Nick Levene ⁶², Lynn Monaghan ⁶², Alexander J Keeley ⁹³, David G Partridge ^{78, 93}, Mohammad Raza ^{78, 93}, Cariad Evans ^{78, 93} and Kate Johnson ^{78, 93}.

Sequencing and analysis:

Emma Betteridge ⁹⁹, Ben W Farr ⁹⁹, Scott Goodwin ⁹⁹, Michael A Quail ⁹⁹, Carol Scott ⁹⁹, Lesley Shirley ⁹⁹, Scott AJ Thurston ⁹⁹, Diana Rajan ⁹⁹, Iraad F Bronner ⁹⁹, Louise Aigrain ⁹⁹, Nicholas M Redshaw ⁹⁹, Stefanie V Lensing ⁹⁹, Shane McCarthy ⁹⁹, Alex Makunin ⁹⁹, Carlos E Balcazar ⁹⁰, Michael D Gallagher ⁹⁰, Kathleen A Williamson ⁹⁰, Thomas D Stanton ⁹⁰, Michelle L Michelsen ⁹¹, Joanna Warwick-Dugdale ⁹¹, Robin Manley ⁹¹, Audrey Farbos ⁹¹, James W Harrison ⁹¹, Christine M Sambles ⁹¹, David J Studholme ⁹¹, Angie Lackenby ⁶⁶, Tamyo Mbisa ⁶⁶, Steven Platt ⁶⁶, Shahjahan Miah ⁶⁶, David Bibby ⁶⁶, Carmen Manso ⁶⁶, Jonathan Hubb ⁶⁶, Gavin Dabrera ⁶⁶, Mary Ramsay ⁶⁶, Daniel Bradshaw ⁶⁶, Ulf Schaefer ⁶⁶, Natalie Groves ⁶⁶, Eileen Gallagher ⁶⁶, David Lee ⁶⁶, David Williams ⁶⁶, Nicholas Ellaby ⁶⁶, Hassan Hartman ⁶⁶, Nikos Manesis ⁶⁶, Vineet Patel ⁶⁶, Juan Ledesma ⁶⁷, Katherine A Twohig ⁶⁷, Elias Allara ^{64, 88}, Clare Pearson ^{64, 88}, Jeffrey K. J. Cheng ⁹⁴, Hannah E. Bridgewater ⁹⁴, Lucy R. Frost ⁹⁴, Grace Taylor-Joyce ⁹⁴, Paul E Brown ⁹⁴, Lily Tong ⁴⁸, Alice Broos ⁴⁸, Daniel Mair ⁴⁸, Jenna Nichols ⁴⁸, Stephen N Carmichael ⁴⁸, Katherine L Smollett ⁴⁰, Kyriaki Nomikou ⁴⁸, Elihu Aranday-Cortes ⁴⁸, Natasha Johnson ⁴⁸, Seema Nickbakhsh ^{48, 68}, Edith E Vamos ⁹², Margaret Hughes ⁹², Lucille Rainbow ⁹², Richard Eccles ⁹², Charlotte Nelson ⁹², Mark Whitehead ⁹², Richard Gregory ⁹², Matthew Gemmell ⁹², Claudia Wierzbicki ⁹², Hermione J Webster ⁹², Chloe L Fisher ²⁸, Adrian W Signell ²⁰, Gilberto Betancor ²⁰, Harry D Wilson ²⁰, Gaia Nebbia ¹², Flavia Flaviani ³¹, Alberto C Cerda ⁹⁶, Tammy V Merrill ⁹⁶, Rebekah E Wilson ⁹⁶, Marius Cotic ⁸², Nadua Bayzid ⁸², Thomas Thompson ⁷², Erwan Acheson ⁷², Steven Rushton ⁵¹, Sarah O'Brien ⁵¹, David J Baker ⁷⁰, Steven Rudder ⁷⁰, Alp Aydin ⁷⁰, Fei Sang ¹⁸, Johnny Debebe ¹⁸, Sarah Francois ²³, Tetyana I Vasylyeva ²³, Marina Escalera Zamudio ²³, Bernardo Gutierrez ²³, Angela Marchbank ¹⁰, Joshua Maksimovic ⁹, Karla Spellman ⁹, Kathryn McCluggage ⁹, Mari Morgan ⁶⁹, Robert Beer ⁹, Safiah Afifi ⁹, Trudy Workman ¹⁰, William Fuller ¹⁰, Catherine Bresner ¹⁰, Adrienn Angyal ⁹³, Luke R Green ⁹³, Paul J Parsons ⁹³, Rachel M Tucker ⁹³, Rebecca Brown ⁹³ and Max Whiteley ⁹³.

Software and analysis tools:

James Bonfield ⁹⁹, Christoph Puethe ⁹⁹, Andrew Whitwham ⁹⁹, Jennifer Liddle ⁹⁹, Will Rowe ⁴¹, Igor Siveroni ³⁹, Thanh Le-Viet ⁷⁰ and Amy Gaskin ⁶⁹.

Visualisation:

Rob Johnson ³⁹.

1 Barking, Havering and Redbridge University Hospitals NHS Trust, **2** Basingstoke Hospital, **3** Belfast Health & Social Care Trust, **4** Betsi Cadwaladr University Health Board, **5** Big Data Institute, Nuffield Department of Medicine, University of Oxford, **6** Brighton and Sussex University Hospitals NHS Trust, **7** Cambridge Stem Cell Institute, University of Cambridge, **8** Cambridge University Hospitals NHS Foundation Trust, **9** Cardiff and Vale University Health Board, **10** Cardiff University, **11** Centre for Clinical Infection & Diagnostics Research, St. Thomas' Hospital and Kings College London, **12** Centre for Clinical Infection and Diagnostics Research, Department of Infectious Diseases, Guy's and St Thomas' NHS Foundation Trust, **13** Centre for Enzyme Innovation, University of Portsmouth (PORT), **14** Centre for Genomic Pathogen Surveillance, University of Oxford, **15** Clinical Microbiology Department, Queens

Medical Centre, **16** Clinical Microbiology, University Hospitals of Leicester NHS Trust, **17** County Durham and Darlington NHS Foundation Trust, **18** Deep Seq, School of Life Sciences, Queens Medical Centre, University of Nottingham, **19** Department of Infection Biology, Faculty of Infectious & Tropical Diseases, London School of Hygiene & Tropical Medicine, **20** Department of Infectious Diseases, King's College London, **21** Department of Microbiology, Kettering General Hospital, **22** Departments of Infectious Diseases and Microbiology, Cambridge University Hospitals NHS Foundation Trust; Cambridge, UK, **23** Department of Zoology, University of Oxford, **24** Division of Virology, Department of Pathology, University of Cambridge, **25** East Kent Hospitals University NHS Foundation Trust, **26** East Suffolk and North Essex NHS Foundation Trust, **27** Gateshead Health NHS Foundation Trust, **28** Genomics Innovation Unit, Guy's and St. Thomas' NHS Foundation Trust, **29** Gloucestershire Hospitals NHS Foundation Trust, **30** Great Ormond Street Hospital for Children NHS Foundation Trust, **31** Guy's and St. Thomas' BRC, **32** Guy's and St. Thomas' Hospitals, **33** Hampshire Hospitals NHS Foundation Trust, **34** Health Data Research UK Cambridge, **35** Health Services Laboratories, **36** Heartlands Hospital, Birmingham, **37** Hub for Biotechnology in the Built Environment, Northumbria University, **38** Imperial College Hospitals NHS Trust, **39** Imperial College London, **40** Institute of Biodiversity, Animal Health & Comparative Medicine, **41** Institute of Microbiology and Infection, University of Birmingham, **42** King's College London, **43** Liverpool Clinical Laboratories, **44** Maidstone and Tunbridge Wells NHS Trust, **45** Manchester University NHS Foundation Trust, **46** Microbiology Department, Wye Valley NHS Trust, Hereford, **47** MRC Biostatistics Unit, University of Cambridge, **48** MRC-University of Glasgow Centre for Virus Research, **49** National Infection Service, PHE and Leeds Teaching Hospitals Trust, **50** Newcastle Hospitals NHS Foundation Trust, **51** Newcastle University, **52** NHS Greater Glasgow and Clyde, **53** NHS Lothian, **54** Norfolk and Norwich University Hospital, **55** Norfolk County Council, **56** North Cumbria Integrated Care NHS Foundation Trust, **57** North Tees and Hartlepool NHS Foundation Trust, **58** Northumbria University, **59** Oxford University Hospitals NHS Foundation Trust, **60** PathLinks, Northern Lincolnshire & Goole NHS Foundation Trust, **61** Portsmouth Hospitals University NHS Trust, **62** Princess Alexandra Hospital Microbiology Dept., **63** Public Health Agency, **64** Public Health England, **65** Public Health England, Clinical Microbiology and Public Health Laboratory, Cambridge, UK, **66** Public Health England, Colindale, **67** Public Health England, Colindale, **68** Public Health Scotland, **69** Public Health Wales NHS Trust, **70** Quadram Institute Bioscience, **71** Queen Elizabeth Hospital, **72** Queen's University Belfast, **73** Royal Devon and Exeter NHS Foundation Trust, **74** Royal Free NHS Trust, **75** Sandwell and West Birmingham NHS Trust, **76** School of Biological Sciences, University of Portsmouth (PORT), **77** School of Pharmacy and Biomedical Sciences, University of Portsmouth (PORT), **78** Sheffield Teaching Hospitals, **79** South Tees Hospitals NHS Foundation Trust, **80** Swansea University, **81** University Hospitals Southampton NHS Foundation Trust, **82** University College London, **83** University Hospital Southampton NHS Foundation Trust, **84** University Hospitals Coventry and Warwickshire, **85** University of Birmingham, **86** University of Birmingham Turnkey Laboratory, **87** University of Brighton, **88** University of Cambridge, **89** University of East Anglia, **90** University of Edinburgh, **91** University of Exeter, **92** University of Liverpool, **93** University of Sheffield, **94** University of Warwick, **95** University of Cambridge, **96** Viapath, Guy's and St Thomas' NHS Foundation Trust, and King's College Hospital NHS Foundation Trust, **97** Virology, School of Life Sciences, Queens Medical Centre, University of Nottingham, **98** Wellcome Centre for Human Genetics, Nuffield Department of Medicine, University of Oxford, **99** Wellcome Sanger Institute, **100** West of Scotland Specialist Virology Centre, NHS Greater Glasgow and Clyde, **101** Department of Medicine, University of Cambridge, **102** Ministry of Health, Sri Lanka, **103** NIHR Health Protection Research Unit in HCAI and AMR, Imperial College London, **104** North West London Pathology, **105** NU-OMICS, Northumbria University, **106** University of Kent, **107** University of Oxford, **108** University of Southampton, **109** University of Southampton School of Health Sciences, **110** University of Southampton School of Medicine, **111** University of Surrey, **112** Warwick Medical School and Institute of Precision Diagnostics, Pathology, UHCW NHS Trust, **113** Wellcome Africa Health Research Institute Durban and **114** Wellcome Genome Campus.

